

THE CHARM-01 STUDY:
ASSESSING
FORMULATIONS OF
TENOFIVIR 1% GEL IN HIV
SERONEGATIVE ADULTS
VIA TRANSCRIPTOME
ANALYSIS

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CHARM-01

- Double-blinded, randomized, safety & acceptability, pharmacokinetic, and ex vivo efficacy study of rectally-applied tenofovir-based microbicide formulations.
- Dose comparison of 3 current formulations of tenofovir 1% gel
 - Vaginal formulation
 - Reduced glycerin formulation
 - Rectal specific formulation
- Endpoints
 - General and mucosal safety
 - Pharmacokinetics and pharmacodynamics
- Current status
 - Completed

Transcriptomic Study Objective and Aims

Transcriptome: The complete set of coding and non-coding transcripts in a given sample and their quantity

Expanded Objective: Apply low-input RNA-Seq transcriptional analysis as a sensitive assay to uncover changes to the mucosal environment caused by gel usage.

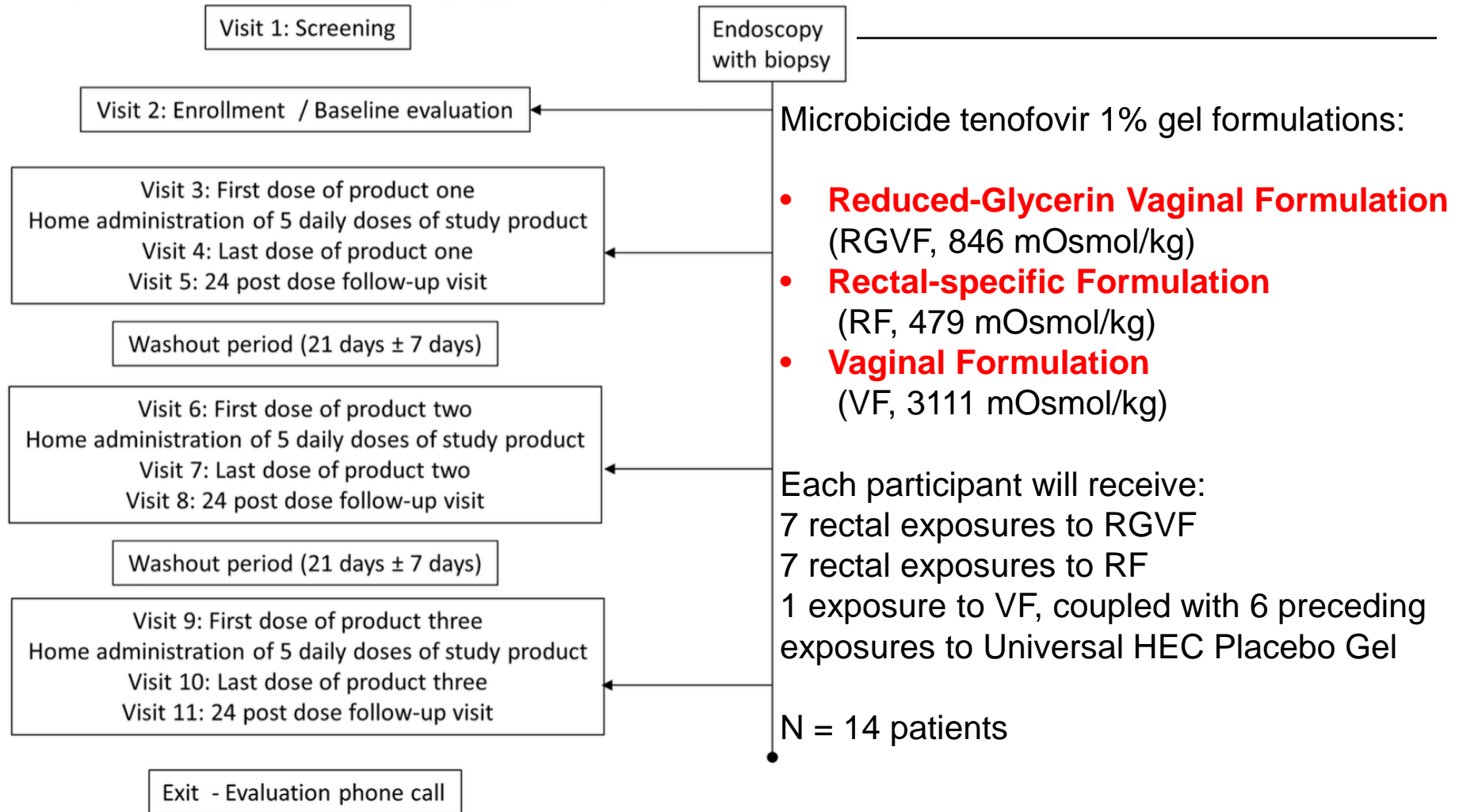
Hypothesis: Local changes in the mucosal immune environment (e.g. IFNs/inflammasome) may be part-and parcel the action of microbicide treatment and alter risk of HIV infection

Aim1: To distinguish the effects of three microbicide tenofovir 1% gel formulations, administered rectally, using transcriptomics

Aim2: To compare data with other microbicide trials and validate signatures of intestinal mucosal gene expression following microbicide exposure associated with protection or risk

CHARM-01 Study Design

Blinded crossover design with Tenofovir 1% formulations in random sequence



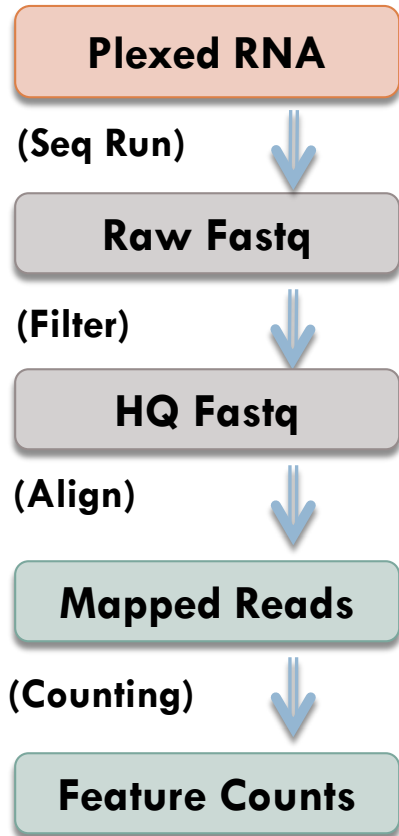
Sample Processing

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- Total RNA was isolated from 50 gut biopsies preserved in *RNAlater* using Qiagen RNEasy Mini Plus Kits
- RNA-Seq data was generated using Illumina Truseq (low input SOP) kits and the Illumina HiSeq 2500
 - ▣ Paired-end, 50 cycle, $>30 \times 10^6$ mapped reads/sample
 - ▣ Medium depth: Able to measure the transcriptome and common splicing variants.
 - ▣ RNA-Seq's advantage areas over microarray include sensitivity, specificity and quantification, as well as fielding coding (mRNA) and non-coding (e.g. small RNA) transcript counts

Preprocessing of RNA-seq: R Pipeline

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1. Create **Raw fastq** from binary output by running pooled tagged RNA **short read libraries** are run on Illumina Hiseq 2500 (2x50 30M reads), as per manufacturer's instructions.
2. Create **High Quality Fastq's** by **filtering out poor quality base calls and adapter contamination** using *Trimmomatic*.
3. Generate **Mapped Read** files by aligning the HQ Fastq reads using the *STAR* aligner.
4. Generate **Features Counts** using *Htseq*.

Gene based Analysis

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Counts

(Import)



Raw Gene Exprs

(QC)



Gene Exprs

(Model)



DE Gene Lists

(Pathway
Enrichment)

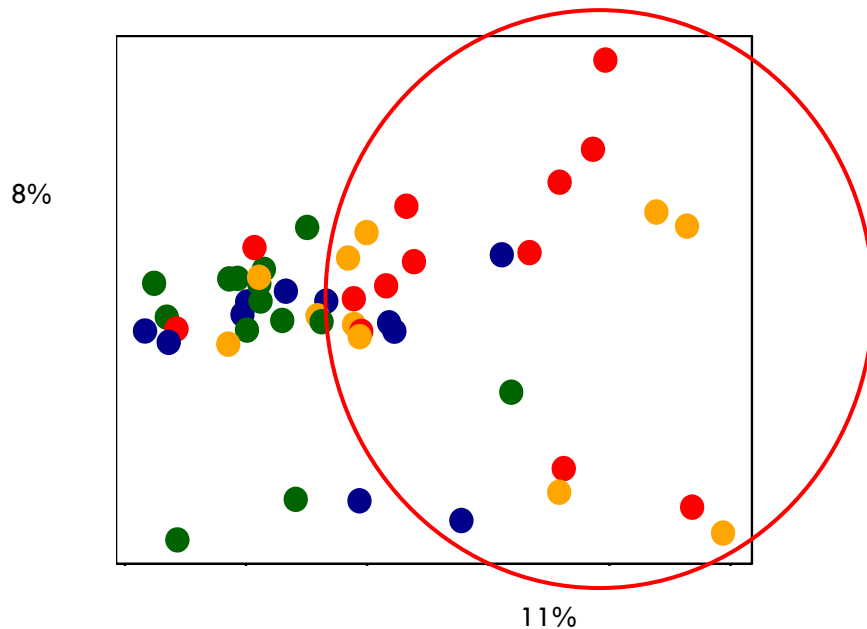


Pathway Lists

1. Create **Raw Gene Expression Matrix (Genes by Samples)** from aggregating count files into a matrix, and importing the sample **Phenotype** data in R.
2. Create **Normalized Expression Matrix** by performing removing sample outliers base on QC assessment and **normalizing the samples to each other** using *Edge*.
3. Generate **Differential Gene Expression Lists** by performing 2 group analysis via linear modeling using *EdgeR*.
4. Generate **Pathway Enrichment Lists** by taking top ranking genes and performing pathway enrichment using Gene Set Enrichment Analysis or *GSEA*.

Effects of three microbicide tenofovir 1% gel formulations on intestinal mucosal gene expression

Comparing formulations to baseline: F-Test



Kruskal wallis rank sum test

Dimension 1

Group	p-value
Baseline	0.002
RGVF	0.001

RGVF and Baseline significantly explain the first dimension of variation

Comparing (Formulation VS Baseline)

Method : Fitting GLM (GLM : R - EdgeR)

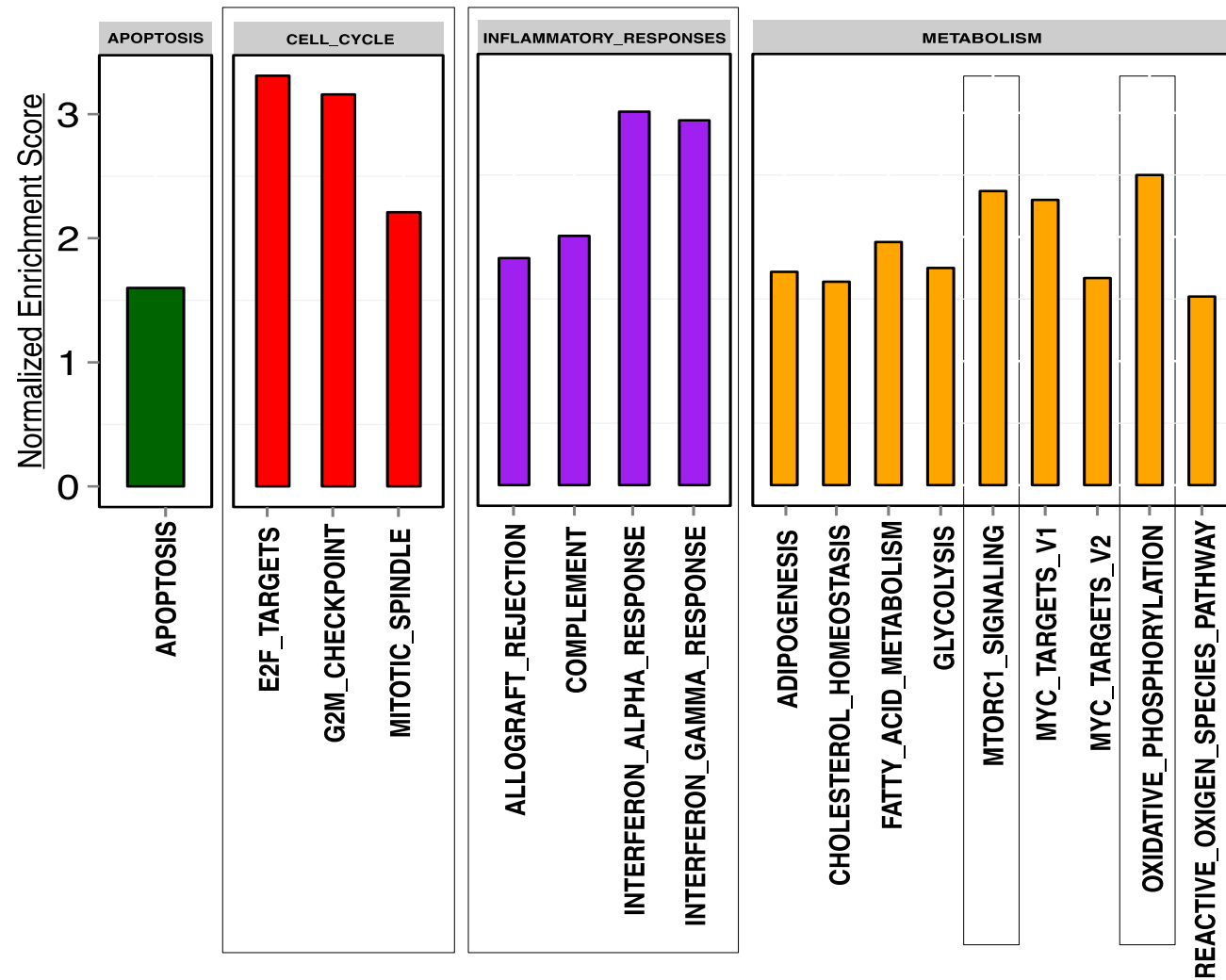
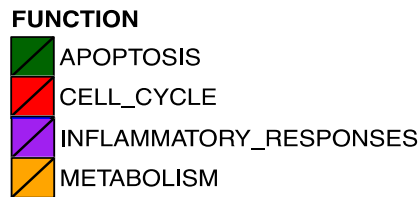
Model : (Gene expression ~ Formulation group + Donor)

Multi Dimensional Scaling analysis of the top 500 F-test genes by P value shows **RGVF has the most significant expression profile compared to **baseline****

Comments: There is a lot of variability between donors (n=12-14) in gene expression observed within the formulation groups.

Solution: Gene Set Enrichment Analysis : RGVF – Baseline Focused

Many increased pathway activities upon exposure to RGVF

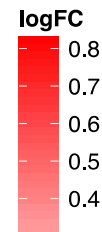
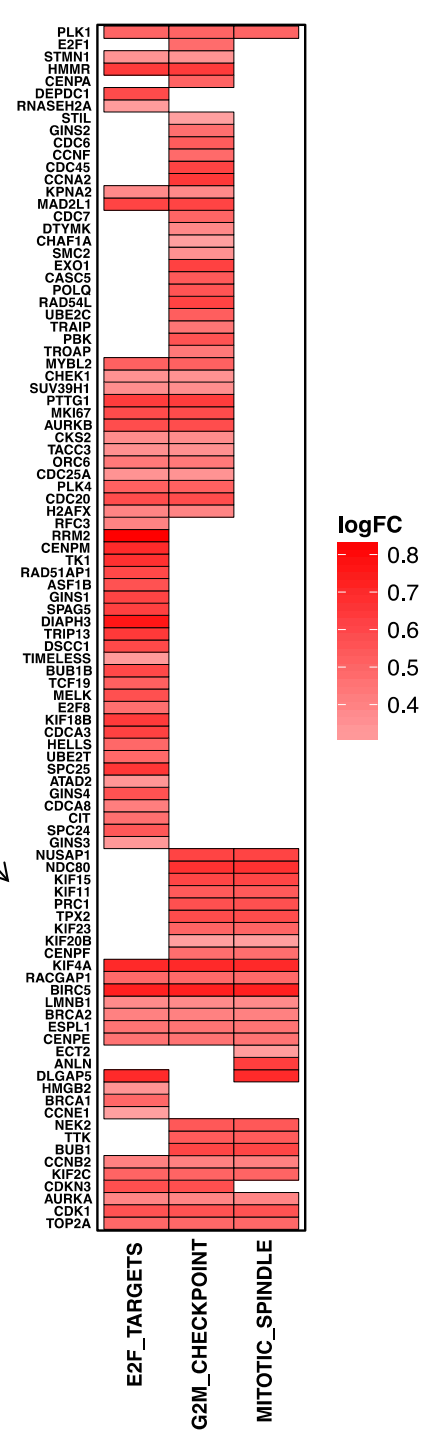
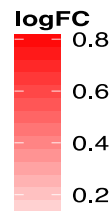
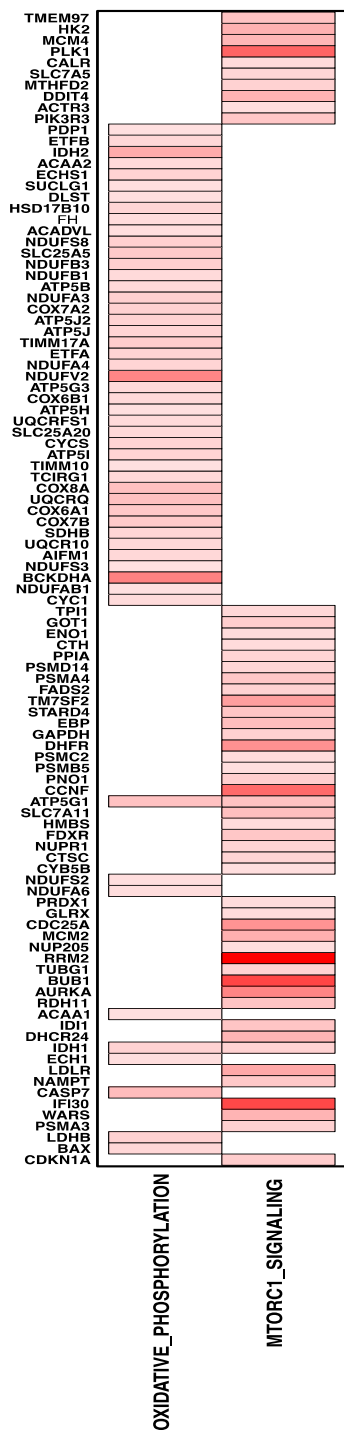


Oxidative phosphorylation, mTORC1 signaling, And cell cycle genes are up-regulated upon exposure to RGVF formulation

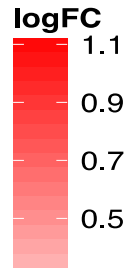
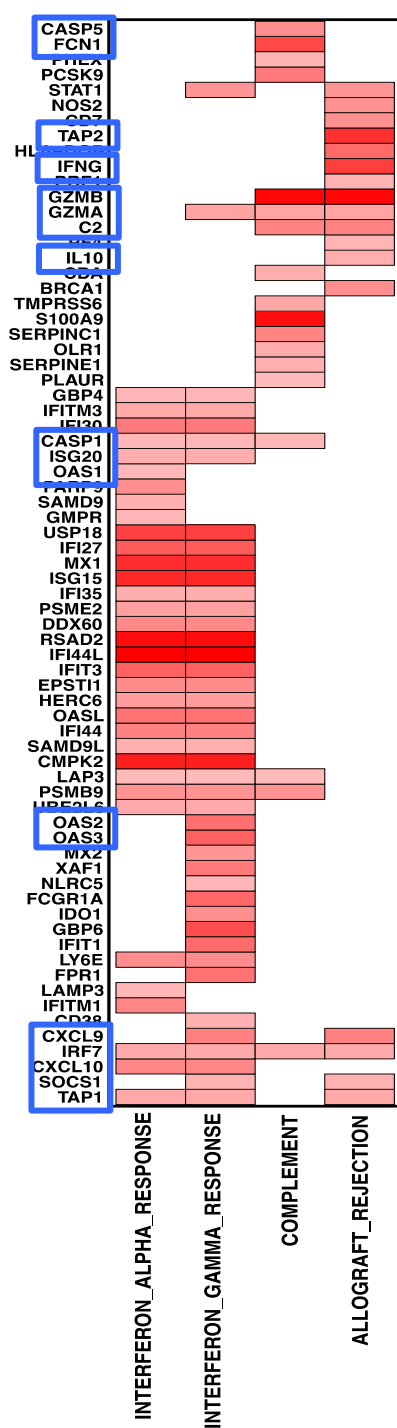
- Genes encoding proteins involved in oxidative phosphorylation
- Genes encoding cell cycle and related targets of E2F transcription factors
- Genes important for mitotic spindle assembly
- Impact on cell proliferation, stress response

Ph related?

- Activation of the mTORC1 complex
- Impact on senescence and T cell fate



Inflammasome-related pathways are up-regulated upon exposure to RGVF formulation

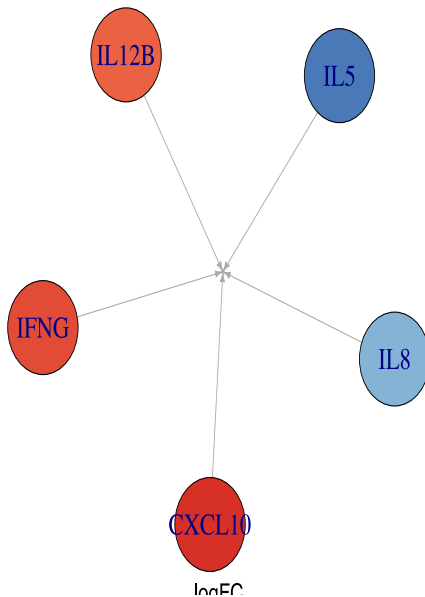


- Genes up-regulated in response to IFNA
E.g. OAS1, LAMP3, IFITM1
- Genes up-regulated in response to IFNG
E.g. CXCL9, SOCS1, GZMA
- General IFN response genes
E.g. IRF7, CXCL10, IFIs/IFITs, MX1, ISG20
- Genes associated with the complement system
E.g. SERPINs, FCN1, C2, CASP1/5
- Genes up-regulated during allograft rejection
E.g. TAP1/2, IL10, IFNG, HLAs

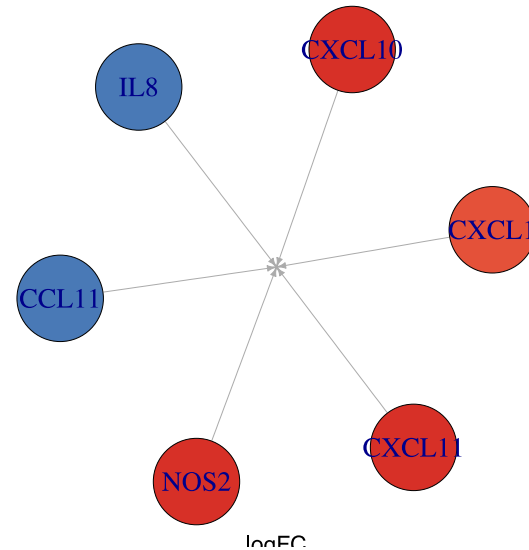
Comparison: Project Gel GSEA: (RG Tenofovir Day1)

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Communication Between
Innate and Adaptive
Immunity

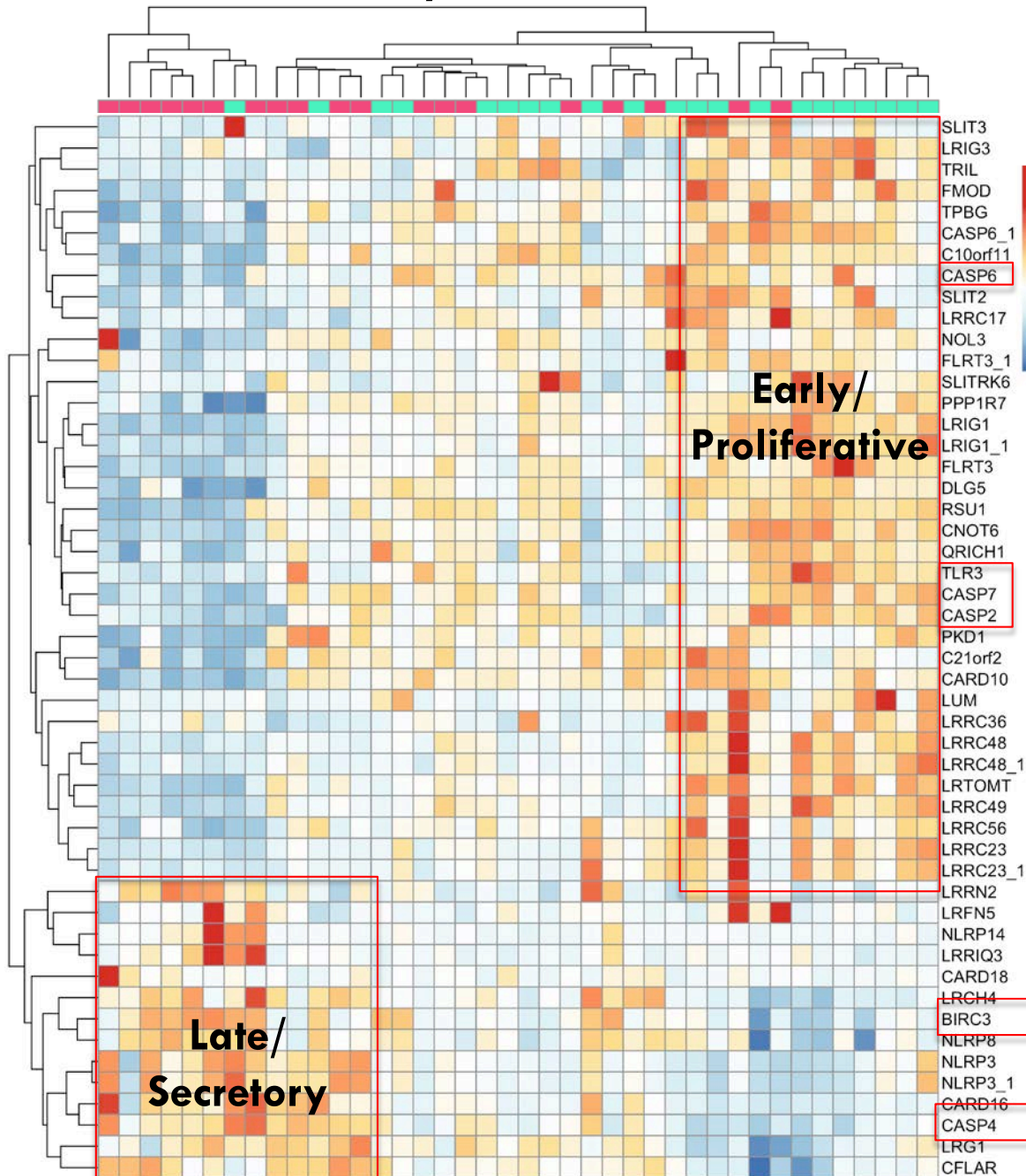


Chemokine Signaling



IFNG-mediated pathway signatures do not persist to D7 in Project Gel, however...

Cytobrushes

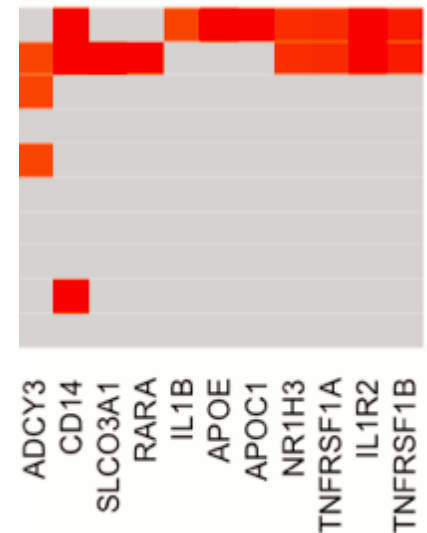


Inflammasome signatures are associated with early/late menstrual cycle phases

Inflammasome-filtered
(Gaucher et al., 2008)

Subset
■ Cytobrush_Early
■ Cytobrush_Late

CVL Samples at Early Phase

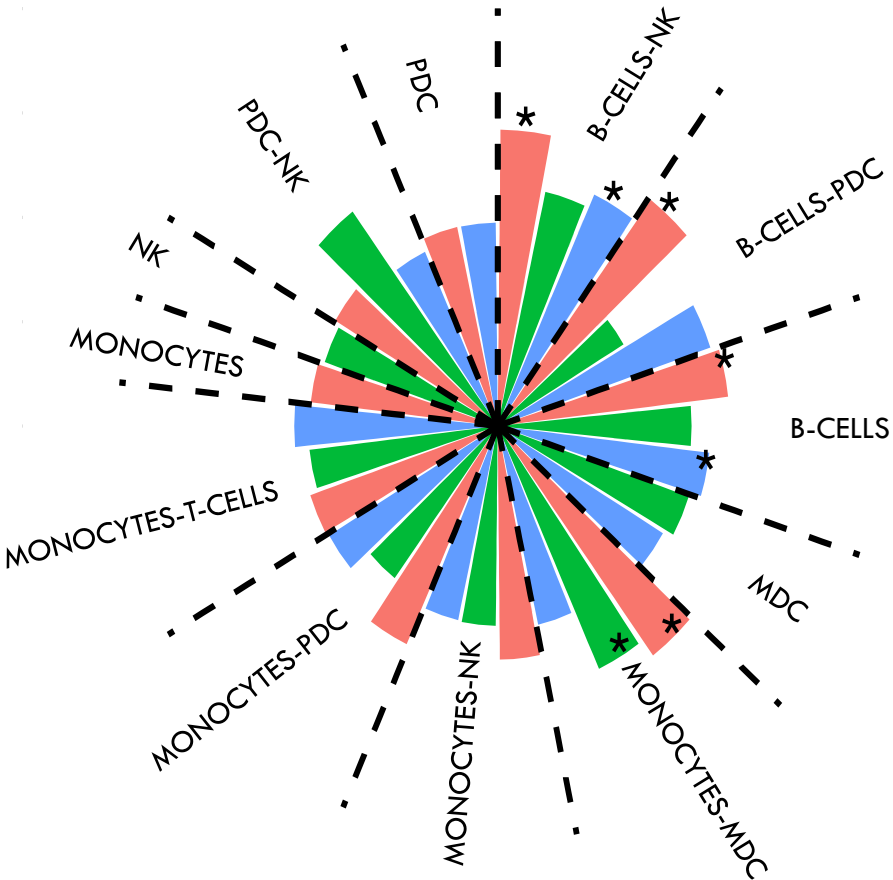


BIRC3: regulates caspase cascades inflammatory gene signaling

Enrichment in B cell and monocyte-related genes describes the D1 and D7 Project Gel gene expression signature



Deconvolution: OMIC Module-Based Filtering
 Nakaya et al. (*Nat Immunol* **12**(8): 786, 2011)



virConst_Day7vsPreExposure

* Pvalue < 0.05

Conclusions:

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- While the N and effect sizes are small, we have begun to hone a *signature* associated with rectal application of tenofovir gel
- RGVF has the most unique expression profile compared to baseline
 - ▣ Increased pathway activities include proinflammatory pathways, IFNs, cell signaling, stress and cell cycle.
 - ▣ Coordinated inflammasome involvement in RGVF formulation
 - ▣ Balance between antiviral IFNs and the greater inflammasome is likely delicate in determining an outcome and a result of many factors
 - ▣ Need outcomes to give this balance context in risk
- CHARM inflammation signature can be seen early in Project Gel
 - ▣ IFNG/IFNG-response genes, SERPINS, SOCS, chemokines are up-regulated
- Nakaya filtering (a deconvolution method) indicate that gene activity in Project Gel may stem from alterations in B cell and Monocytes/MDC.

A lot of work remains:

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Perform additional contrasts to pair an individual subject with their baseline and formulation sequence and map their overall responses in the MDS and inflammasome

- hampered by lower subject numbers and resulting bioinformatic approaches (e.g. ranking DGE by nominal P)

Increase subject numbers, integrate different data types (flow data, proteome) and *outcomes*, train and test signatures in similar human trials, or meta-analysis across very different studies (human, NHP)

Develop and validate new flow panel and/or Fluidigm PCR panel to probe these signatures at a finely sorted and in titration down to the single cell level

Identify and target mechanisms and biomarkers of protection or infection risk in future trials

Acknowledgements

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